CLEVEACEAE-MARCHANTIALES

SAUTERIA NYIKAENSIS, A NEW LIVERWORT SPECIES FROM MALAWI

INTRODUCTION

In the Cleveaceae the dorsal air pores of the thalli are simple and the radial walls of the surrounding cells are often thickened. Three genera have traditionally been grouped together in this family, namely *Athalamia* Falc., *Peltolepis* Lindb. and *Sauteria* Nees. *Peltolepis* and *Sauteria* have not been reported from Africa, but *Athalamia* (formerly *Clevea*) has long been known from this continent, with two species that occur here, namely A. *spathysii* (Lindenb.) Nees and A. *pulcherrima* (Steph.) Hatt. (Vanden Berghen 1965). A third species, *Clevea* (Athalamia) crassa Trabut, from the Atlas Mountains (Magreb), is considered to be a nom. inval. (Grolle 1976).

Sauteria is a small genus of \pm five (Bischler 1998) or six species (Schuster 1992), although some authorities recognize only three species worldwide (Gradstein *et al.* 2001). The

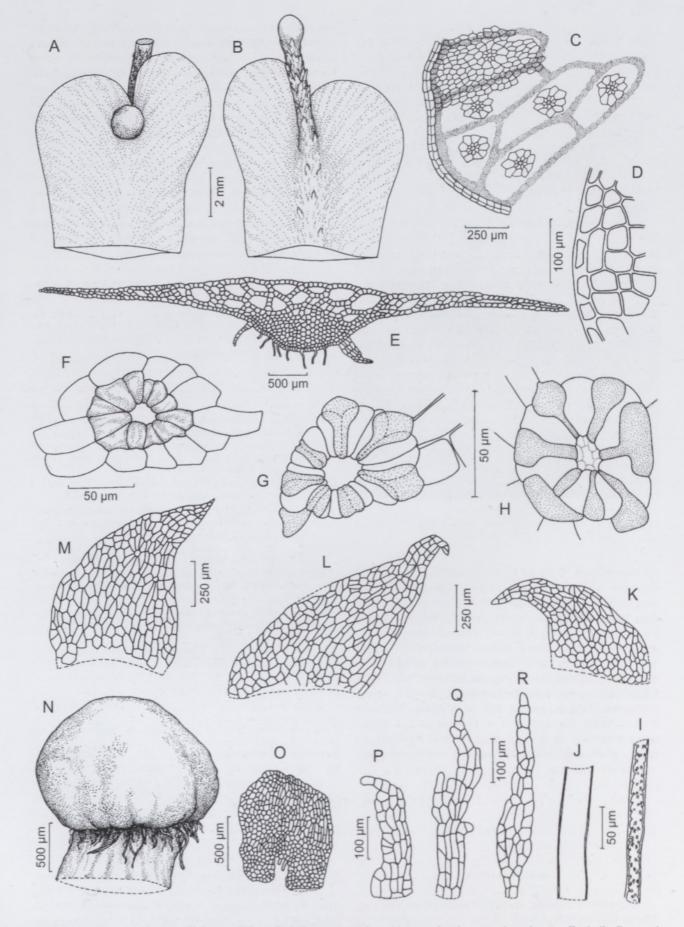


FIGURE 6.—Sauteria nyikaensis, Koekemoer 1874. A–E, thallus: A, dorsal face with young female receptacle, stolon cut off apically; B, ventral face with stolon uncut, covered with scales and rhizoids; C, air chambers partly exposed beneath dorsal epidermal cells and air pores surrounded by thickened cells; D, marginal cells and some dorsal epidermal cells; E, c/s. F–H, air pore and surrounding cells from above. I, J, rhizoids: I, tuberculate; J, smooth-walled. K–M, ventral scales; N, young female receptacle from side; O, c/s stalk with single rhizoid furrow; P–R, paleae. Drawings by M. Steyn.

genus is widespread, but is restricted to high mountain environments, often above 3 000 m, although it also occurs at lower elevations on islands (Gradstein *et al.* 2001). In Europe, Russia, Siberia, Tibet, the Himalayas and northern Japan, the distribution of its members is arctic or montane. Its western range includes areas in Iceland, east and west Greenland, as well as east and west Canada (Müller 1951–1958). Hitherto, the only known records in the southern hemisphere are those from the Andes of Peru, northern Argentina and Chile, as well as the Galapagos Islands, where it was found at an altitude of about 1 200 m (Gradstein *et al.* 2001).

During April 2000, an unusual thallose liverwort was collected on the Nyika Plateau at an altitude of 2 343 m, on soil, in a cavity under a rock overhang. Unfortunately, neither of the two female receptacles present is mature, and antheridia are absent. Nevertheless, in order to draw attention to this plant, it is described here and has been referred to the genus Sauteria for the following reasons: 1, the thalli are light green and fragile, the assimilation tissue is spongy and there is no trace of pigmentation; 2, the cells surrounding the simple air pores are strongly thickened; 3, the air chambers are empty, lacking both filaments and papillae, medianly in 2 or 3 layers and, visible beneath the epidermal cells in the wings, are the parallel outlines of what appears to be a single layer, obliquely orientated toward the thallus margins; 4, the ventral scales do not extend to the thallus margins; they are hyaline, with a single, tapering appendage, and are arranged in ill-defined rows, mostly confined to the prominent midrib; oil bodies are very rare and apical slime papillae are absent; 5, the female receptacle originates from a deep notch at the apex of the thallus; 6, in cross section the stalk of the receptacle has a single rhizoid furrow, whereas Athalamia species have none and Peltolepis species have two.

According to Schuster (1992), 'Sauteria is separated from the other two genera of the Cleveaceae by one absolute feature (solitary rhizoid furrow of carpocephalum stalk)'. He also mentions 'distinct, scattered oil cells' in the ventral scales, in some (but not all) cases, adding that isolated ventral scales in Greenland Sauteria alpina often lack oil cells. Shimizu & Hattori (1954) describe the oil cells in S. alpina as 'scattered in the ventral scales and the dorsal epidermis of thallus, rare'. In their description of S. alpina var. japonica (later elevated to S. yatsuensis), they note that, 'oil-cells scattered in ventral tissue of thallus and bractlets of female receptacle (and also in ventral scales of thallus?), very rare', their question mark clearly indicating uncertainty. In a later description of S. alpina, Hattori & Shimizu (1955) remark that 'oil-cells rare, scattered in ventral tissue, ventral scales and bractlets'. They do not, however, illustrate oil cells in their 'Text-fig. XXI', although figs F-H of the ventral scales show groups of 5-7 cells surrounding a much smaller central cell, which does not contain an oil body. This is also seen in Figure 6K-M of the Nyika plant. Oil bodies in Sauteria have been observed to be long-persisting; those in the scales of S. alpina, leg. S.O.Lindberg & E.Rettig (held at PRE), are still present 120 years after collection.

Sauteria nyikaensis Perold, sp. nov.

Thalli magnitudine media vel sat magna, apice semel dichotome ramificantes, interdum irregulariter; laete virides, sine pigmento, fragiles spongiosique. Cavernulae aeriae, circumscriptione clare visibile, in medio thalli parallele, apicem versus dispositae, sed marginem thalli versus oblique dispositae. Pori dorsales non elevati, simplices, ab 1 vel 2 annulis concentricis cellularum circumscripti, pro parte vel pro parte maxima incrassationibus conspicuis tecti. Squamae ventrales hyalinae appendiculo acuminato non semper manifesto, in seriebus incertis supra costam dispositae. Costa saepe producta stolonem magnum geotropicum formans. Antheridia non visa. Receptaculum gynoeciale immaturum, in incisura apicali setae brevi tereti, sulco uno rhizoidali insidens. Guttae olei omnino absentes, semel tantum in squama ventrali visae.

TYPE.—Malawi, 1033: Nyika National Park, Jalawe viewpoint, (-BD), on soil, in a cavity under rock overhang, at altitude 2 343 m, 3 April 2000, *Koekemoer 1874* (PRE, holo.) with *Lunularia cruciata* (L.) Dumort. ex Lindb. and *Plagiochasma eximium* (Schiffn.) Steph.

Thalli prostrate, in crowded patches, obovate, apically notched or incised, on either side with rounded lobes (Figure 6A, B), continuing sometimes as smaller lobes along slightly decurved (Figure 7A) attenuate margins; medium-sized to fairly large, up to 12 mm long and 5-9 mm wide distally, narrowing gradually or abruptly up to ±4 mm wide proximally, branching dichotomously once, but in young plants often irregularly or rather diffusely; light green, without any pigmentation, margins colourless; fragile and spongy, with clearly visible outlines of elongated, empty air chambers medianly running parallel toward apex, but soon becoming obliquely orientated toward thallus margins (Figure 6C), each one apparently opening by a simple air pore; along dorsal midline, slightly concave and not grooved, laterally margins acute, flanks sloping obliquely, ventral face medianly keeled with a prominent midrib, rounded distally but flattening proximally, covered with rhizoids and illdefined rows of hyaline scales; midrib rarely branched at its apex, ensuing laminae irregularly shaped, most commonly continuing growth distally and occasionally proximally as well, forming very large tuberous, geotropic stolons (Figures 6AB; 7D), up to 850 µm diam., filled with starch grains. Dried plants with flanks sometimes flat, incurved or ascending.

Dorsal epidermal cells rarely chlorophyllose, unistratose, thin-walled, without trigones, 4- to 6-sided, shorter than wide, generally 30-45 × 60.0-72.5 µm, in cross section 32.5-40.0 µm thick; margins unistratose, with 2 juxtaposed cell rows (Figure 6D), mostly rectangular, others 5-sided, outermost cells 22.5-50.0 × 15.0-32.5 µm, some with thickened walls between adjoining cells; second row of cells 4- or 5-sided, 22.5-45.0 × 27.5-45.0 µm, walls not thickened; air pores (Figures 6F-H; 7B; 8A) one per air chamber, not raised, simple, oval or rounded, $10-15 \times 10-20 \ \mu m$, with or without faint inner ring of small cells and then bounded by 1, occasionally 2 concentric rings of cells, variable in number, and covered partly to sometimes almost entirely by conspicuous thickenings, 15-25 × 12.5-20.0 µm, also obscuring several of the radial cell walls, width of air pore together with surrounding cells 62.5-100.0 µm; row of dorsal epidermal cells adjoining thickened cells often

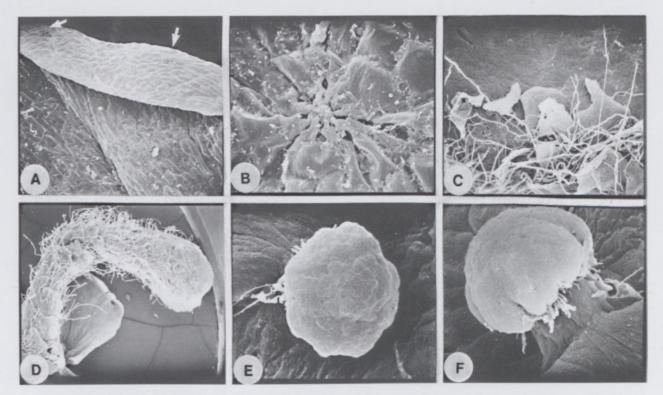


FIGURE 7.—Sauteria nyikaensis, Koekemoer 1874. SEM micrographs. A, margin of dorsal face of thallus decurved over lateral part of ventral face, arrows indicating positions of some dorsal air pores with thickenings; B, thickened cells around contracted air pore in concentric rings, partly shown; C, irregular rows of ventral scales mostly over midrib, between rhizoids; D, massive ventral stolon with scales and rhizoids, partly overlying apical part of ventral face of thallus; E, young female receptacle from above; F, young female receptacle from side. A, × 58; B, × 430; C, × 18.5; D, × 8.5; E, × 18; F, × 21.

somewhat smaller than average, partly arranged in a concentric ring, occasionally the thickenings extending slightly onto a few of them as well. (Figure 8A, B), $22.5-25.0 \times 20.0-27.5 \,\mu$ m, light brown and finely granular.

Assimilation tissue, as seen below and through dorsal epidermis, with parallel outlines of empty air chambers 200-350 µm apart, running obliquely across wings to margins of thallus, partitioned in wings by slanting, unistratose cross walls at intervals of up to 600 µm between them, at thallus margins air chambers somewhat smaller; in cross section (Figure 6E), thallus over midrib 700-1150 µm thick, ± upper half with polyhedral air chambers in 2 or 3 layers, $65-175 \times 150-220 \,\mu\text{m}$, with lower ones smaller, unistratose walls consisting of chlorophyllose cells, spherical or ovoid, $37.5-50.0 \times 25.0-42.5 \,\mu\text{m}$; storage tissue occupying $\pm \frac{1}{6}$ of width of thallus medianly and ± 12 rows of cells in lower half of thickness of thallus, decreasing laterally, soon disappearing and flanks bounded beneath by ventral epidermis only, cells crowded together, angular, 27.5-40.0 × 50-65 um, no sclerotic cells, oil bodies or mucilage cavities present; rhizoids densely covering midrib, fewer beneath wings, some smooth (Figure 6J), 27.0-47.5 wide, others pegged (Figure 6I), 15-25 µm wide. Ventral scales (Figure 6K-M), hyaline, in 2-4 poorly defined rows over midrib and extending onto adjacent ventral face of thallus (Figure 7C), also on continuation(s) of midrib as geotropic stolon(s) (Figures 6B; 7D); inconspicuous, asymmetrically triangular, one side obliquely rounded, margins entire, tapering gradually and not constricted where joined with acuminate, apically pointed and not sharply differentiated appendage, 725-1025 µm long (including appendage), width across base 375-725 µm, cells 4-6-sided, 45-75 × 30-45 µm, in each scale 1-3 groups of cells surrounding 1 much smaller, central cell, not containing an oil body; oil bodies very rare

Monoicous? Antheridia unknown. Gynoecial receptacle terminal, raised on short stalk, (Figures 6N; 7E, F) at crotch of apical incision up to 2.5 mm long, separating 2 thallus lobes, immature, rounded above, ± 1675 µm wide, with 8 lobes below, air pores not seen, but may develop later, as air chambers visible in cross section of receptacle, a single archegonium also seen; stalk terete, with one rhizoid furrow (Figure 6O), 625 µm long at this stage of development, 875 µm wide, without assimilatory strip, naked below, but with paleae at apex; paleae elongated and narrow (Figures 6P-R; 9B) 375-800 × 75-120 µm, inner cells 25-40 × 12.5-25.0 µm, with 1 to 3 papillae apically and sometimes 1 at margin, thickerwalled at tip. Chromosome number for the genus Sauteria: n = 36 (Müller 1951–1958; Hattori & Shimizu 1955, count by Dr S. Tatuno for Sauchia japonica, later transferred to Sauteria yatsuensis; Bischler 1998). The Nyika material was no longer living when examined, and a chromosome count could not be done.

DISCUSSION

In the absence of antheridia, the single archegonium seen in the above specimen, would not have been fertilized. The development of antheridia may have been delayed for some reason, or else they had already disappeared. Bischler (1998) states that in families of the Marchantiales with archegoniophores other than the Marchantiaceae, the stalk elongates after fertilization. In the Nyika plant, however, the stalk is still very short, almost sessile, and fertilization had not taken place.

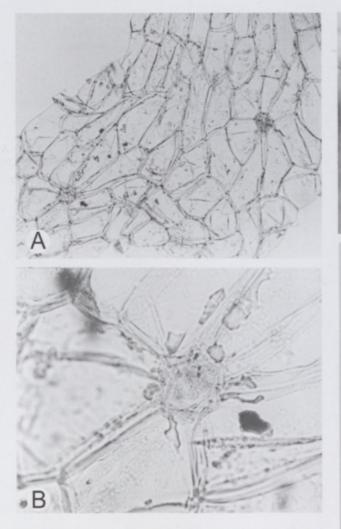


FIGURE 8.—Sauteria nyikaensis, Koekemoer 1874. LM micrographs. A, part of ventral scale with 2 oil bodies; B, much enlarged micrograph of one of the oil bodies. A, × 198; B, × 790.

Because of the delicate structure of the thalli, the plants are thought to be drought intolerant and to perennate during the dry season by means of the tuberous geotropic stolons.

At the site in northern Malawi (Figure 10), where the plant was collected near the Jalawi View Point, it grew on calcareous soil containing slivers of mica, in a small, cave-like cavity at the base of a large rock. Not much direct sunlight could have reached it there, but species of Sauteria, except for S. chilensis, lack secondary pigmentation even when growing in open, strongly illuminated sites (Schuster 1992). Shimizu & Hattori (1954) do not regard the presence of thickened radial walls around the dorsal air pores as being of generic value, since they had observed pores with both thickened and thin radial walls on the same plant. All the air pores in the Nyika specimen had thickenings, not just on the radial walls but partly or entirely covering the cells surrounding them, somewhat like those in Athalamia pulcherrima, as illustrated by Vanden Berghen (1965). When stained with periodic acid-Schiff's (PAS) reaction (Jensen 1962), the thickenings became intensely pink, much more so than the rest of the tissues.

B FIGURE 9.—Sauteria nyikaensis, Koekemoer 1874. LM micrographs. A, dorsal air pore with thickenings on surrounding cells; B, palea. A, × 500; B, × 100. FIGURE 9.—Sauteria nyikaensis, Koekemoer 1874. LM micrographs. A, dorsal air pore with thickenings on surrounding cells; B, palea. A, × 500; B, × 100. same family, the Cleveaceae, but adds that, 'other characters argue against such a classification'. She does not elaborate further. Sauteria nyikaensis is separated from the other species in the genus by collectively considering the following characters listed in Table 1.

The genus *Sauteria* was first described by Nees (1838) and named for the Austrian physician, Anton E. Sauter, 1800–1881, who also collected and studied liverworts.

If more material of *Sauteria nyikaensis* with ripe sporophytes is collected, the above description will, undoubtedly, have to be emended.

ACKNOWLEDGEMENTS

Bischler (1998) states that the genera Athalamia, Sauteria and Peltolepis are traditionally grouped in the Koekemoer, for collecting this specimen while on a South-



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	S. alpina (Schuster 1992)	S. chilensis (Hässel de Menéndez 1963)	S. nyikaensis (Perold 2003)	S. yatsuensis (Hattori & Shimizu 1955)
Distribution	Central and northern Europe, Yugoslavia, Siberia, Norway, Sweden, Finland, Iceland, Greenland, Alaska, British Columbia, Alberta	Andes of Peru, northern Argentina and Chile; Galapagos Islands	Nyika Nat. Park, Malawi	Mt Rishiri, Rishiri Island, Hokkaido, northern Japan
Thallus				
colour	light green, margins decolorate	dark green, margins & ventral face purple	light green, no pigmentation	pale to light or dull green
size	$8-17 \times 3-5 \text{ mm}$	$6-12 \times 1.5-2.5 \text{ mm}$	up to $12 \times 5-9$ mm distally	$10-15 \times (3-)5-7 \text{ mm}$
branching	simple or $1-2 \times dichotomous$	$3-4 \times dichotomous$	dichotomous or irregular	sparsely dichotomous
segments	lingulate	branches with apical innovations	rounded lobes	
apex	deeply emarginate	furrowed	notched or incised	
texture	fleshy, ± soft		fragile, spongy	'not so firm'
margins	thin, translucent, ascending	purple	colourless, 2 juxtaposed cells unistratose, attenuate	
dorsal epidermis	strongly areolate	cells with chloroplasts	unistratose, cells thin-walled	
pores	± elevated, bounded by 5–7 cells, radial walls thickened	not raised, bounded by 5 or 6 cells, radial walls thickened	not raised, bounded by up to 12 or 13 cells, mostly covered by thickenings	radial walls of cells not so thickened, often thin and indistinct
air chambers	inflated, polyhedral, (2)3(4) layers	1(2) layer(s), narrow-rectangular	polyhedral, 2 or 3 layers	polygonal
Ventral scales	not projecting at thallus margins	extending to thallus margins	over midrib and occasionally on adjacent ventral face	hardly reaching thallus margins
rows	irregular, 3-6	in 2 irregular rows	2-4 poorly defined rows	in 3-5(6) irregular rows
colour	hyaline	reddish black or decolorate	hyaline	colourless
shape	asymmetrically ovate-lanceolate	base triangular	asymmetrically triangular	ovate or lanceolate
appendage	acuminate to longly acute	ciliate, basally constricted or not	acuminate, not sharply differentiated	narrowed to 1-celled apex
marginal slime papillae present, clavate	lae present, clavate	ephemeral	absent	usually present
oil bodies	scattered in scale body	in 1 or 2 isolated cells	very rare	rare
Geotropic stolons	not mentioned	not mentioned	present, prominent	not mentioned
Androecia	ill-defined group behind female stalk	behind female receptacle	not seen	just below female receptacle or on different branch
Gynoecia	from deep apical notch	at apical bifurcation	from deep apical notch	disc non-convex, lacking pores
stalk	colourless, up to 15 mm long	yellow-green, 2-8 mm long	immature, only 625 µm long	
naleae	none at base. few at anex		narrow and elongated, at apex	

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FIGURE 10.-Locality of Sauteria nyikaensis in Malawi.

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REFERENCES

- BISCHLER, H. 1998. Systematics and evolution of the genera of the Marchantiales. *Bryophytorum Bibliotheca* 51: 1–201.
- EDWARDS, D. & LEISTNER, O.A. 1971. A degree reference system for citing biological records in southern Africa. *Mitteilungen der Botanischen Staatssammlung München* 10: 501–509.
- GAO, C., ZHANG, G.C. & CAO, T. 1981. Taxa nova bryophytorum tibeticanum. Acta Botanica Yunnanica 3: 389–399.
- GRADSTEIN, S.R., CHURCHILL, S.P. & SALAZAR-ALLEN, N. 2001. Guide to the bryophytes of tropical America. *Memoirs of the New York Botanical Garden* 86: 1–577.

GROLLE, R. 1976. Verzeichnis der Lebermoose Europas und benachbarter Gebiete. Feddes Repertorium 87: 171–279.

- HÄSSEL DE MENÉNDEZ, G.G. 1963. Estudio de las Anthocerotales y Marchantiales de la Argentina. *Opera Lilloana* 7: 1–279.
- HATTORI, S. & SHIMIZU, D. 1955. Marchantiales of Japan IV. Journal of the Hattori Botanical Laboratory 14: 91–107.
- JENSEN, W.A. 1962. Botanical histochemistry: 1–408. Freeman, San Francisco & London.
- KASHYAP, S.R. 1929. Liverworts of the western Himalayas and the Panjab Plains. 1: 1–129. Lahore.
- MÜLLER, K. (Müll.Frib.) 1951–1958. Die Lebermoose Europas. Dr. L. Rabenhorst's Kryptogamen-Flora 6, edn. 3: 368–382.
- NEES AB ESENBECK, C.G. 1838. Naturgeschichte der europäischen Lebermoose 4: 1–540.
- SCHUSTER, R.M. 1992. The Hepaticae and Anthocerotae of North America 6: 1–937. Field Museum of Natural History, Chicago.
- SHIMIZU, D. & HATTORI, S. 1954. Marchantiales of Japan. III. Journal of the Hattori Botanical Laboratory 12: 53–75.

VANDEN BERGHEN, C. 1965. Hépatiques récoltées par le Dr J.-J. Symoens dans la région péri-Tanganyikaises. Bulletin de la Société Royale de Botanique de Belgique 98: 129–174.

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