A floristic classification of the vegetation of a forest-savanna boundary in southeastern Zimbabwe

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Keywords: classification, Detrended Correspondence Analysis, forest-savanna boundary, moist forest, savanna, TWINSPAN, vegetation type, Zimbabwe

ABSTRACT

The vegetation of Chirinda Forest boundary was classified into eight types using Two-way Indicator Species Analysis (TWINSPAN) and Detrended Correspondence Analysis (DCA). The moist forest comprises three types: *Strychnos mello-dora–Chrysophyllum gorungosanum* Forest on deep dolerite soils; *Chrysophyllum gorungosanum–Myrianthus holstii* Forest on shallow dolerite soils; and *Teclea nobilis–Ehretia cymosa* Forest on drier, but deep dolerite soils. The non-forest vegetation comprises five types: *Themeda triandra* Grassland on shallow dolerite soils; *Psidium guajava* Bushland on sandstone; *Bridelia micrantha–Harungana madagascariensis* Mixed Woodland not restricted to any one particular soil type; *Acacia karroo–Heteropyxis dehniae* Woodland on shallow soils derived from sandstone but sometimes on dolerite; and *Julbernardia globi-flora–Brachystegia spiciformis* (Miombo) Woodland on sandstone.

INTRODUCTION

Forest patches occur in many tropical countries, often embedded in a matrix of non-forest vegetation types, forming unique units along their boundaries. Ranney *et al.* (1981) emphasized the importance of forest fringes in the structure and dynamics of forest patches. The boundary creates microclimatic gradients that result in differences in environment between the forest interior and the outside, non-forest areas. In this context, the influence of soil moisture and other edaphic changes at the forest-savanna boundary is particularly important (Furley 1992; Hopkins 1992).

The ecotone between forest and non-forest areas often has high plant densities and diversity, and represents the juxtaposition of two contrasting habitats. Along the ecotone there is generally a high incidence of wind and animal activity, especially mammals and frugivorous birds. As a result, the predation of seed and fruit and the opportunity for their dispersal are both enhanced.

In Zimbabwe, forest patches with sharp boundaries between them and non-forest vegetation are common along the eastern highlands where three main centres of moist forest development can be identified, namely the Nyanga, the Vumba and the Chimanimani massifs. Many other smaller patches of forest occur elsewhere, including Chirinda, the area of this study. The factors responsible for these boundaries vary from place to place, but fire, altitude and edaphic factors are among the most important. Crook (1952) and Phipps & Goodier (1962) discussed the vegetation patterns of the Chimanimani Mountains, with emphasis on the determinants of the forest, woodland, grassland and Ericaceous scrub. Similar situations can also be encountered in the Vumba, Nyanga and Chirinda areas. Inventories of the vegetation of Chirinda Forest have been compiled by, among others, Goldsmith (1976) and Muller (1991). Much of this work focused mainly on the moist forest proper, with very little attention paid to the surrounding non-forest vegetation. Mapaure (1993) discussed the factors influencing the vegetation of the forest boundary whilst Timberlake *et al.* (1994) described the composition of the forest and some ecological factors affecting the vegetation. A list of flowering plants and ferns found in and around the forest has been compiled by Drummond & Mapaure (1994).

The main objective of this study was to inventory the woody vegetation of Chirinda Forest boundary, including adjacent bushland, grassland and woodland areas and to produce a classification and description of the vegetation types.

STUDY AREA

The study was carried out in Chirinda Forest, situated near Mount Selinda Mission (20° 25'S, 32° 43'E) in southeast Zimbabwe (Figure 1). The forest represents the southern end of moist forest distribution in Zimbabwe and is the best preserved example of medium altitude moist forest (Muller 1991). It is surrounded by pine plantations, Mission settlements, Chako Business Centre, and commercial farmlands. It is a gazetted Forest Land managed by the Forestry Commission, covering 947 ha, of which 606 ha is moist evergreen forest. The forest lies on two broad highlands rising from 1 076 m to 1 250 m in altitude. These form two crests, essentially subdividing the forest into two, a northern and a southern section with a narrow forested saddle in between.

The geology of the area comprises red and purple phyllitic mudstones interbedded with pale fine- to mediumgrained feldspathic sandstones of the Upper Argillaceous series of the Precambrian Umkondo System (Watson 1969). Fine-grained dolerite sills have intruded into these sandstones, forming a cap over much of the higher

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ground. The moist forest is generally found on dolerite soils.

The area receives an average rainfall of 1 470 mm per annum (Anon. 1977). Average winter and summer temperatures are 14°C and 20°C, respectively (Sayce 1987). The prevailing winds are southeasterly.

METHODS

An interpretation of 1:12 500, black-and-white aerial photographs of 1987 was carried out and stratification of the vegetation around the forest was done, based on the textural differences on the photographs. A map was produced based on both the interpretation of the aerial photographs, which aided in the marking of the boundaries of the vegetation types, and field sampling.

Ten belt transects (two through each of the five vegetation units apparent on the aerial photographs), each 200 m long and 20 m wide, were established (Figure 2A–J) so that half the distance was in the forest and the other half in the non-forest vegetation. The edge of the forest was identified by either an obvious noticeable change in species composition or by a difference in vegetation structure, or both. Each belt transect was subdivided into ten 20×20 m contiguous quadrats, five on either side of the forest boundary, resulting in a total of 100 quadrats. The appropriateness of this quadrat size was confirmed by a species-area curve.



In each quadrat, the woody species were identified and assigned to height classes as follows: seedlings, saplings (< 0.5 m), understorey (0.5-3.0 m), subcanopy trees (3-10 m), and canopy trees (>10 m). An overall cover-abundance value for each woody species in each quadrat was assessed following the Braun-Blanquet scale (Mueller-Dombois & Ellenberg 1974). Climbers were rated separately according to the number of stems encountered in the stands rather than for a cover value.

Soil depth was determined by augering six quadrats per transect, three on either side of the forest edge. The soils were considered deep if no rocks were encountered down to a depth of 80 cm. The geology was determined from any exposed bedrock and by reference to the geological map of the area.

Data analysis

Vegetation data were analysed using Two Way Indicator Species Analysis (TWINSPAN) (Hill 1979) and Detrended Correspondence Analysis (DCA) (Gauch 1982). TWINSPAN was applied on the full species data set, consisting of 261 plant species belonging to 204 genera and 76 families and DCA was applied on the sixty quadrats from which soil data were collected. Rare species were downweighted and for pseudospecies cut-levels of 0, 1, 3 and 5 were used in the application of TWINSPAN. Coverabundance values of species were used in both analyses. Minor refinements were done to the TWINSPAN output (Table 1) to improve its clarity, especially the removal, after analysis, of the species which occurred three times or less (except indicator species) and lianes and vines.

RESULTS AND INTERPRETATION

Vegetation classification

Eight vegetation types were identified (Figures 3 & 4) on the basis of the TWINSPAN and DCA analyses. The primary division of the stands by TWINSPAN separated out the moist forest stands from non-forest (woodland and grassland) stands. The second level of division separated out forest stands in the northwestern part of the forest from the rest of the forest stands. These forest stands (Type III) are situated close to the miombo stands, on the drier side of the highland. The third level of division further subdivided the remaining forest stands into a group which occurred in the eastern to southeastern part of the forest (mainly transects E, F, G and H) (Type II). The other group (Type I) consists mostly of forest stands from transects A, B, C and D. The forest stands, therefore, fall into three main types.

Among the non-forest vegetation stands, the second level of division separated out grassland stands, which occurred in the southeastern part of the forest (Type VII), from the rest of the non-forest vegetation. The remaining non-forest stands were divided into a further two groups at the third level of division. One type (Type IV), which consists of a wide mixture of stands from several transects but close to the apparent forest edge, was separated from the rest of the remaining non-forest vegetation. The remaining stands were further divided into two, one of which comprises two types. Type VI consisted of stands mainly from an area close to Chako Business Centre (the sandstone enclave) and stands from the northwestern side forming Types V and VIII. Further subdivision of these types was considered unnecessary because the number of stands in each group was too small to justify the subdivision.

The forest, woodland and grassland types were clearly separate, with some ubiquitous intermediary vegetation types bridging them. Within the three moist forest vegetation types, two were not clearly separated on the basis of species composition but were strongly differentiated on



FIGURE 2.—A map showing the major vegetation types in Chirinda Forest area. The positions of the transects (A–J) are indicated.

the relative dominance of the species in the upper canopy and subcanopy strata. *Strychnos mellodora, Rothmannia urcelliformis*, and *Chrysophyllum gorungosanum* had higher cover abundance values (averaging about 50%) in forest Type I than in forest Type II, where they averaged 20%. *Myrianthus holstii* had a lower cover value (about 10%) in forest Type I than in Type II (about 25%). Forest Type III occurred in a drier area, had a more open canopy and,



FIGURE 3.—Dendrogram showing the TWINSPAN results of the quadrats in Chirinda Forest (eigenvalues are shown in brackets).

TABLE 1.—A synoptic table of TWINSPAN classification of Chirinda Forest vegetation. Full names of species indicated by mnemonics are given in Appendix 1

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TABLE 1.—A synoptic table of TWINSPAN classification of Chirinda Forest vegetation. Full names of species indicated by mnemonics are given in Appendix 1 (continued)

		Stand number									
		2223347111237 1 78908090890987897	34444555655666677 76789067689078907	1 899908891237 078908906666	5 1177889 5645346764	2223333344445 53451234523454	67266677999 55223412345	688888992 112345121	111 123123	4555 1123	
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hence, its species composition and undergrowth development were somewhat different from the other two forest types. *Dracaena fragrans* was the most abundant shrub species with an average cover abundance value of more than 50% in forest Type II, especially where the soils were shallow and the terrain steep. The vegetation types are described below. *Strychnos mellodora–Chrysophyllum gorungosanum* Forest (Type I)

This forest consists of a lower upper canopy of various species with occasional emergents. The average height is generally lower (about 30–40 m) than the other forest types (which are about 50–60 m in most areas) except for



FIGURE 4.—A DCA ordination diagram showing the grouping of stands.

190

the emergents. It is dominated by *Strychnos mellodora*, *Chrysophyllum gorungosanum*, *Craibia brevicaudata* and *Tannodia swynnertonii*. Emergent species include *Strychnos mitis*, *S. usambarensis* and strangler *Ficus* spp., in particular *F. chirindensis*. The lower layer (which is not markedly different from the upper canopy) is dominated by *Strychnos mellodora*, *Rawsonia lucida*, *Heinsenia diervilleoides* and *Tabernaemontana ventricosa*. This forest type is found mainly in the area surrounding the sandstone enclave. Common shrubs include the young of the canopy species and low densities of *Dracaena fragrans*. *Acacia pentagona* and *Landolphia buchananii* are the most common lianes.

Chrysophyllum gorungosanum–Myrianthus holstii Forest (Type II)

This forest is dominated by *Chrysophyllum gorungosanum*, *Myrianthus holstii*, *Strombosia scheffleri* and *Craibia brevicaudata* in the upper canopy stratum. The area covered by this type coincides with the wetter parts of the forest. *Khaya anthotheca* is quite common in some places. *Dracaena fragrans* is the dominant shrub and apparently locally excludes other shrubs due to its high densities. *Strychnos mellodora* is largely absent from the subcanopy, which is instead dominated by *Heinsenia diervilleoides*, *Rothmannia urcelliformis*, *Tannodia swynnertonii* and *Tabernaemontana ventricosa*. *Cola greenwayi* and *Drypetes gerrardii* are also fairly common. The most common lianes are *Acacia pentagona*, *Hippocratea goetzei*, *H. pallens* and *Oncinotis tenuiloba*.

Teclea nobilis-Ehretia cymosa Forest (Type III)

This forest consists of a mixture of species of apparently drier forest affinities. It is dominated by *Teclea nobilis* and *Ehretia cymosa*. Common associates include *Chrysophyllum gorungosanum, Diospyros abyssinica* and *Ochna arborea*. It is found in the northwestern part of the forest adjacent to an area which seems to have been cultivated during the 1940s. *Celtis africana, Newtonia buchananii* and *Peddiea africana* form the dominant group in the shrublayer. There is high prevalence of small trees of pioneer species such as *Newtonia buchananii* and *Croton sylvaticus*.

Bridelia micrantha-Harungana madagascariensis Mixed Woodland (Type IV)

This woodland, which appears to be more of a secondary nature, is more developed in the area just south of the Mount Selinda Mission on a north-facing slope, where it forms a belt extending towards the southeastern part of the forest boundary. It is characterized by large, but widely spaced, Bridelia micrantha trees with occasional Macaranga capensis and Cussonia spicata. The lower stratum is dominated by Harungana madagascariensis. Thickets of Toddalia asiatica, Asparagus falcatus and, sometimes, Smilax anceps are a common feature of the shrub layer. Harungana madagascariensis, however, ceases to be dominant in some localities where Pterocarpus rotundifolius trees and Albizia gummifera saplings form an important component of the woodland. Peddiea africana shrubs and Croton sylvaticus seedlings are also common.

Acacia karroo-Heteropyxis dehniae Woodland (Type V)

This woodland, which is dominated by Acacia karroo, Heteropyxis dehniae and Faurea saligna, is on the drier side of the forest and just merges into the miombo woodland (dominated by Julbernardia globiflora, Uapaca kirkiana, U. sansibarica and Brachystegia spiciformis). It is mostly found between the moist forest proper and welldefined non-forest associations. It contains a wide variety of species including pioneer species such as Bridelia micrantha, Albizia gummifera, Croton sylvaticus and Newtonia buchananii. A form of this vegetation type also occurs in small patches elsewhere around the forest, with varying degrees of dominance among the component species. Occasional emergents include Parinari curatellifolia, Catha edulis and Primus africana. Shrubs are represented by Psidium guajava, Vangueria apiculata, Rhus longipes and R. transvaalensis.

Psidium guajava Bushland (Type VI)

This bushland is dominated by the exotic *Psidium guajava* and by *Canthium mundianum* bushes and sometimes assumes the structure of a low woodland. It is found in the sandstone enclave. A few emergent trees of *Parinari curatellifolia* and *Albizia gummifera* occur. Thickets of *Toddalia asiatica, Lantana camara* and *Rubus rigidus* are common, particularly near the boundary with the moist forest. *Peddiea africana* and *Pteridium aquilinum* are also common.

Themeda triandra Grassland (Type VII)

This bushed grassland vegetation is dominated by *Themeda triandra* and *Cymbopogon caesius*. It occurs on shallow dolerite soils with occasional exposed boulders. There is local variation in grass species dominance, with *Loudetia simplex* apparently becoming more dominant on east-facing slopes of the southern part of the forest where the soils are deeper and mostly sandy, sometimes with *Parinari curatellifolia* bushes. Other woody species found within the *Themeda* grassland include *Psidium guajava*, *Eriosema nutans, Pseudarthria hookeri* and *Athrixia rosmarinifolia*.

Julbernardia globiflora–Brachystegia spiciformis Woodland (Type VIII)

An additional woodland type to those apparent in the TWINSPAN classification was identified by DCA, a miombo woodland dominated by Julbernardia globiflora and Brachystegia spiciformis with Uapaca kirkiana, Heteropyxis dehniae and Faurea saligna as common associates in the canopy layer. It is relatively open, with poorly developed shrub and herb layers. Common trees in the subcanopy layer include Heteropyxis dehniae, Faurea saligna, Canthium mundianum, Julbernardia globiflora and Brachystegia spiciformis. Grasses include Digitaria gazensis and Themeda triandra. The TWINSPAN analysis lumps this type with Acacia karroo-Heteropyxis dehniae Woodland (Type V) but DCA clearly separates it from the rest.

Ordination

The first DCA axis, which accounted for 63% of the observed variation, separated the forest and adjacent nonforest vegetation types, with a wide gap of up to one DCA unit between them (Figure 4). In general, stands close to the forest edge, from either side, were closer to the centre of Axis 1 than stands further from the forest edge. Nevertheless, the stands lying adjacent to one another along the forest edge were quite separate with 0.5–2 DCA units between them. Three groups of forest stands were distinguishable, corresponding to those identified in the classification by TWINSPAN. These groups are not separated along the first DCA axis but are distinguishable along the second DCA axis, which accounted for 19.6% of the variation. A similar pattern is also apparent in the ordination of the woodland groups of stands.

Miombo woodland is clearly separated from other woodland stands along the second DCA axis. The grassland stands are, likewise, clearly separated from the woodland stands. There is considerable overlap between the stands from the sandstone enclave and those from the northwestern part of the forest (i.e. the *Psidium guajava* Bushland (Type VI) and *Acacia karroo-Heteropyxis dehniae* Woodland (Type V).

The existence of the Chrysophyllum gorungosanum-Myrianthus holstii Forest (Type II) on the southeastern to the southwestern sides adjacent to the grassland was associated with a higher altitude and a wetter moisture regime, regardless of the shallow soils. Psidium guajava Bushland (Type VI) occurred where the soils were predominantly sandstone and relatively deep. Where the sandstone soils were steeper and shallower, miombo and sometimes Acacia karroo-Heteropyxis dehniae Woodland (Type V) occurred. On dolerite, with the same conditions, Strychnos mellodora-Chrysophyllum gorungosanum Forest (Type I) occurred, whereas the Teclea nobilis-Ehretia cymosa Forest (Type III) occurred where it was drier. The Bridelia micrantha-Harungana madagascariensis Mixed Woodland (Type IV) occurred on predominantly dolerite soils with other factors being intermediate. This was also found in what appeared to be geological transitional zones.

DISCUSSION

Vegetation types

The vegetation of Chirinda Forest boundary has been classified into eight types. This represents only those types occurring within 100 m on either side of the forest edge.

The TWINSPAN analysis clearly separated forest from non-forest stands. The stands close to the forest edge, however, were not clearly separated on the basis of whether they were just inside or just outside the forest, as might have been expected. The apparent lack of clear differentiation may be due to the existence of some species that are transitional between forest and woodland. Such a vegetation unit, which normally consists of both fire-tolerant savanna and fire-tender forest tree species (Hopkins 1992), represents a mosaic of communities of each of the two vegetation types. The transitional vegetation between savanna and forest shows great variation in both structure and species composition from place to place around the forest. Some of the types described in this category are restricted in extent whereas others occur widely, though in patches.

The transitional vegetation includes forest pioneer species. Most pioneer species in Chirinda Forest are Afromontane endemics or near-endemics which are absent or rare in lowland forests. These include Albizia gummifera, Anthocleista grandiflora and Maesa lanceolata. Hence, the forest has sometimes been labelled a 'transitional' forest because of the co-existence of both lowland and Afromontane forest species (White 1978). Even though the majority of the moist forest species have lowland phytogeographic affinities, several species such as Chrysophyllum gorungosanum (which is one of the commonest), Casearia battiscombei, Drypetes gerrardii, Halleria lucida, Myrianthus holstii, Prunus africana, Strombosia scheffleri and Xymalos monospora are Afromontane in origin. Chirinda Forest is, therefore, Afromontane in origin and character but with lowland phytogeographic affinities.

The southern section of the forest supports *Chrysophyllum gorungosanum–Myrianthus holstii* Forest (Type II), with more *Khaya anthotheca* and *Trichilia dregeana*, whereas the northern section mainly supports the *Strychnos mello-dora–Chrysophyllum gorungosanum* Forest (Type I) with more *Craibia brevicaudata* and *Strychnos mitis*. Perhaps the greatest difference between these forest types manifests itself in the variation in species composition of the understorey. This difference seems to be determined by soil depth and slope, which also influence the moisture regime. Shallow and steep sites drain faster than areas of moderate slope, leading to differences in the undergrowth species composition. Understorey species are therefore important in defining the limits of these two types.

The forest and woodland vegetation types share only a few species between them. Greater overlap in species composition was observed between the forest and the ecotonal vegetation than between forest and, for instance, miombo woodland. Shared species appear to occur mostly as seedlings, and less so as trees, in those habitat types to which they do not characteristically belong. Some of these, such as *Croton sylvaticus, Harungana madagascariensis* and *Bridelia micrantha* are, however, true pioneers which thrive well under gap conditions in forests.

Ordination

Forest generally occurs in wetter environments, and savanna in drier ones (Backeus 1992). In Chirinda, the amount of precipitation received from rainfall in the area is lower than the normal requirement for forest development. The extra moisture comes in the form of orographic drizzle, made possible by several factors: the high ground, the southeasterly aspect, and the tall trees which facilitate the release of this extra moisture from the low clouds. The southern section of the forest has shallower soils, but a higher average altitude and receives more moisture from the south-easterlies, advected in from the Mozambique Channel, than the northern section. In general, Chirinda receives up to 28% more moisture than the average of five surrounding stations, much of which can be attributed to orographic drizzle (Mapaure 1993).

Miombo Woodland (Type VIII) was clearly classified separately from the Acacia karroo-Heteropyxis dehniae Woodland (Type V) by DCA but not by TWINSPAN. This might have been due to the apparent importance attached to Heteropyxis dehniae by TWINSPAN in Types V and VIII, resulting in the recognition of these types as one. Also of much interest, is the distance of separation between the dominant forest type [Chrysophyllum gorungosanum-Myrianthus holstii Forest (Type II)] and the rest of the non-forest types on the DCA ordination diagram. The DCA distance between the stands represents the average standard deviation of the species turnover, where a full species turnover occurs in about four DCA units (Gauch 1982). Thus, the DCA units between the types in Chirinda Forest represent about a 50% change in sample species composition, indicating a more or less abrupt change. This change may indicate the strength of geology in determining the extent of the forest, since all forest stands occurred on dolerite.

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APPENDIX 1.—Full names of plants appearing in Table 1. Synonyms in square brackets.

MNEMONIC	FULL NAME AND AUTHORITY	CEL MIL	Celtis mildbraedii Engl.
ACA KAR	Acacia karroo Hayne	CHI BAT	Chionanthus battiscombei (Hutch.) Stearn
ACA SIE	Acacia sieberiana DC.	CHR GOR	Chrysophyllum gorungosanum Engl.
AGE PEN	Agelaea pentaphylla (Lam.) Baill.	CLA ANI	Clausena anisata (Willd.) Hook.f. ex Benth.
ALB GUM	Albizia gummifera (J.FGmel.) C.A.Sm.	CLE SWY	Clerodendrum swynnertonii S.Moore
ANN SEN	Annona senegalensis Pers.	CLU SWY	Clutia swynnertonii S.Moore
ANT VEN	Antidesma venosum Tul.	CNE POL	Cnestis polyphylla Lam.
ARG MAC	Argomuellera macrophylla Pax	COF LIG	Coffea ligustroides S.Moore
ARG TOM	Argyrolobium tomentosum (Andrews) Druce	COL GRE	Cola greenwayi Brenan
ASP PLU	Aspilia pluriseta Schweinf. subsp. pluriseta	COM MOL	Combretum molle R.Br.
ATH ROS	Athrixia rosmarinifolia (Walp.) Oliv. & Hiern	CRA BRE	Craibia brevicaudata (Vatke) Dunn subsp. baptistarum
BEQ NAT	[Bequaertiodendron natalense (Sond.) Heine & J.H.Hemsl.]		(Büttner) J.B.Gillett
	Englerophytum natalense (Sond.) Pennington	CRE TRI	Cremaspora triflora (Thonn.) K.Schum.
BER ABY	Bersama abyssinica Fresen.	CRO SYL	Croton sylvaticus Hochst.
BRA SPI	Brachystegia spiciformis Benth.	CRY LIE	Cryptocarya liebertiana Engl.
BRI MIC	Bridelia micrantha (Hochst.) Baill.	CUS SPI	Cussonia spicata Thunb.
CAL AUR	Calpurnia aurea (Aiton) Benth. subsp. aurea	CYM CAE	Cymbopogon caesius (Hook. & Arn.) Stapf
CAN MUN	Canthium mundianum Cham. & Schltdl.	DES SET	Desmodium setigerum (E.Mey.) Harv.
CAR BIS	Carissa bispinosa (L.) Desf. ex Brenan subsp. bispinosa	DIC CIN	Dichrostachys cinerea (L.) Wight & Arn.
CAS BAT	Casearia battiscombei R.E.Fr.	DID NOR	Didymosalpinx norae Swynn.
CAS MAL	Cassipourea malosana (Baker) Alston	DIO ABY	Diospyros abyssinica (Hiern) F.White
CAT EDU	Catha edulis (Vahl) Endl.	DIO LYC	Diospyros lycioides Desf.
CEL AFR	Celtis africana Burm.f.	DOM BUR	Dombeya burgessiae Harv.
CEL GOM	Celtis gomphophylla Baker	DOV MAC	Dovyalis macrocalyx (Oliv.) Warb.

Bothalia 27,2 (1997)

DRA FRA	Dracaena fragrans Ker Gawl.	PRU AFR	Prunus africana (Hook.f.) Kalkman
DRA MAN	Dracaena mannii Baker	PSE HOO	Pseudarthria hookeri Wight & Arn.
DRY GER	Drypetes gerrardii Hutch.	PSE SUB	Pseuderanthemum subviscosum (C.B.Clarke) Stapf
EHR CYM	Ehretia cymosa Thonn. var. divaricata (Baker) Brenan	PSI GUA	Psidium guajava L.
ERI NUT	Eriosema nutans Schinz	PSO FEB	Psorospermum febrifugum Spach
ERY LYS	Erythrina lysistemon Hutch.	PTE ROT	Pterocarpus rotundifolius (Sond.) Druce
EUC DIV	Euclea divinorum Hiern	RAU CAF	Rauvolfia caffra Sond.
FAD CIE	Fadogia cienkowskii Schweinf.	RAW LUC	Rawsonia lucida Harv. & Sond.
FAU SAL	Faurea saligna Harv.	RHA PRI	Rhamnus prinoides L'Hér.
FIC CHI	Ficus chirindensis C.C.Berg	RHO REV	Rhoicissus revoilii Planch.
FIC SUR	Ficus sur Forssk.	RHU LON	Rhus longipes Engl.
FLE GRA	Flemingia grahamiana Wight & Arn.	RHU TEN	Rhus tenuinervis Engl.
GRE OCC	Grewia occidentalis L. var. occidentalis	RHU TRA	Rhus transvaalensis Engl.
HAL LUC	Halleria lucida L.	RHY SWY	Rhynchosia swynnertonii Baker f.
HAR MAD	Harungana madagascariensis Poir.	RIN FER	Rinorea ferruginea (Baker f.) M.Brandt
HEI DIE	Heinsenia diervilleoides K.Schum.	ROT URC	Rothmannia urcelliformis (Hiern) Bullock
HET DEH	Heteropyxis dehniae Suess.	RUB RIG	Rubus rigidus J.E.Sm.
HET TRI	Heteromorpha trifoliata (H.L. Wendl.) Eckl. & Zevh.	RUB COR	Rubia cordifolia L. subsp. conotricha (Gand.) Verdc.
HYP ARI	Hypoestes aristata (Vahl) Roem. & Schult.	RUT FUS	Rutidea fuscescens Hiern
IND HED	Indigofera hedvantha Eckl & Zevh	SAP ELL	Sapium ellipticum (Hochst.) Pax
IND SWA	Indigofera swaziensis <i>Bolus</i>	SCO STO	Scolonia stolzii <i>Gilg</i>
	Iulbemardia globiflora (<i>Benth</i>) Trounin	SEN LAT	Senecio latifolius DC
KEE GUE	Keetia gueinzii (Sand) Bridson	SEN SEP	Senna septemtrionalis (Viv.) Irwin & Barneby
KHA ANT	Khava anthotheca <i>Baker</i> f	SEN SIN	Senna singueana (Delile) Lock
LAN CAM	Lantana camara I	SPH PRU	Sphedampocarpus pruriens (A Juss) Szyszyl
LIP IAV	Lippia javanica (Burm f.) Spreng	STR MEI	Strychnos mellodora S Maare
LOV SWY	Lovoa swynnertonii Baker f	STR MIL	Strychnos metics S Maare
MAC CAP	Macaranga capensis (Raill.) Sim	STR SPI	Strychnos spinosa Lam
MAELAN	Maesa Janceolata Forssk	STR USA	Strychnos usambarensis Gila
MAY HET	Maximus beterophylla (Fekl & Zevh) N Rubson	STR SCH	Strombosia scheffleri <i>Engl</i>
MAY SEN	Maytenus neterophyna (Lent. & Zeyn.) W.Kooson Maytenus sanagalansis (Lam.) Exall	SVZ GUG	Syzygium guingense (Willd) DC subsp. guingense
MELLOR	Mallera lobulato S Muure	TAR VEN	Tabemaemontana ventricosa Huchst ar ADC
MEL LOB	Murienthus holetii Eugl	TAN SWV	Tannodia suvannertonii (S Maara) Prain
MYK HUM	Myriantnus noistil <i>Engl.</i>	TAD DAV	Tamoula swymertoini (S.Moore) Fram
OCH ARB	Ochna arborea <i>DC</i> . var. oconnorii (<i>E.Phillips</i>) <i>Du Toit</i>	IAK FAV	Bridson
OCI GRA	Ocimum gratissimum L var. gratissimum	TEC NOB	Teclea nobilis Delile
OXY GOE	Oxyanthus goetzei K.Schum. subsp. goetzei	TEP LON	Tephrosia longipes <i>Meisn</i> . subsp. swynnertonii (<i>Baker f.</i>) Brunmitt
OXY SPE	Oxyanthus speciosus DC.	THE TRI	Themeda triandra Forssk.
PAR CUR	Parinari curatellitolia Planch. ex Benth.	TIL FUN	Tiliacora funifera (Miers) Oliv.
PAV COM	Pavetta comostyla S.Moore subsp. comostyla var. comostyla	TRE ORI	Trema orientalis (L.) Blume
PED AFR	Peddiea africana Harv.	TRI DRE	Trichilia dregeana Sond.
PHY NUM	Phyllanthus nummularitolius Poir.	TRI PIN	Triumfetta pilosa Roth var. nyasana Sprague & Hutch.
PLE PYC	Pleiocarpa pycnantha (K.Schum.) Stapf	TRI MAD	Trilepisium madagascariense DC.
POL FUL	Polyscias fulva (Hiern) Harms	VAN API	Vangueria apiculata K.Schum.
PRO FAL	[Protasparagus falcatus (L.) Oberm.] Asparagus falcatus L.	VAN INF	Vangueria infausta Burch.
PRO LAR	[Protasparagus laricinus (Burch.) Oberm.] Asparagus	VER COL	Vernonia colorata (Willd.) Drake
DRO PLU	Internus Durch.	XYL PAR	Xylopia parviflora (A.Rich.) Benth.
TRO TLU	plumosus Baker	XYM MON	Xymalos monospora (Harv.) Baill.