Studies in the Marchantiales (Hepaticae) from southern Africa. 1. The genus *Dumortiera* and *D. hirsuta*; the genus *Lunularia* and *L. cruciata*

S.M. PEROLD*

Keywords: Dumortiera, D. hirsuta, Dumortieroideae, Hepaticae, Lunularia, L. cruciata, Lunulariaceae, Marchantiaceae, Marchantiales, taxonomy, southern Africa, Wiesnerellaceae

ABSTRACT

The genera Dumortiera (Dumortieroideae, Marchantiaceae) and Lunularia (Lunulariaceae), are briefly discussed. Each genus is represented in southern Africa by only one subcosmopolitan species, D. hirsuta (Swartz) Nees and L. cruciata (L.) Dum. ex Lindberg respectively.

UITTREKSEL

Die genusse Dumortiera (Dumortieroideae, Marchantiaceae) en Lunularia (Lunulariaceae) word kortliks bespreek. In suidelike Afrika word elke genus verteenwoordig deur slegs een halfkosmopolitiese spesie, D. hirsuta (Swartz) Nees en L. cruciata (L.) Dum. ex Lindberg onderskeidelik.

DUMORTIERA Nees

Dumortiera Nees ab Esenbeck in Reinwardt, Blume & Nees ab Esenbeck, Hepaticae Javanicae, Nova Acta Academiae Caesareae Leopoldina-Carolinae Germanicae Naturae Curiosorum XII: 410 (1824); Gottsche et al.: 542 (1846); Schiffner: 35 (1893); Stephani: 222 (1899); Sim: 25 (1926); Müller: 394 (1951–1958); S. Arnell: 52 (1963); Hässel de Menéndez: 182 (1963). Type species: Dumortiera hirsuta (Swartz) Nees.

Synonymy according to Nelson & Parnell: 35 (1992):

Hygropyla Taylor: 390 (1836).

Hygrophila Taylor (orth. var.) in Mackay: 53 (1836) non R. Br. (1810).

Hygrophyla Taylor (orth. var.) in Mackay: X (1836). Hygropila Taylor (orth. var.) in J.D. Hooker & T. Taylor: 576 (1844).

Askepos Griffith 2: 340 (1849).

Thallus, large, flat and thin, dark green, in overlying patches; on shaded, damp soil or on wet rocks. Branches with apical innovations or dichotomously furcate, occasionally lateral, moderately divergent, thickened over midrib, wings gradually thinning toward margins; apex emarginate.

Dorsal epidermis very thin-walled, temporary, vestigial, air pores absent, air chambers mostly reduced to 1- or 2-celled vestiges, reported rarely to have reduced chlorophyllose filaments, cells containing numerous fairly large chloroplasts; storage tissue compact, confined to ventral part of midrib; oil cells rare, each with a single, large oil body; rhizoids along midrib and ventrally appressed against wings, mostly smooth, occasionally tuberculate; scales ventral over midrib, vestigial and evanescent. Monoicous or dioicous. Antheridia sunken in subsessile disciform receptacles, which are fringed with bristles and borne singly at apex of thallus on short bifurrowed stalk. Archegonia in groups of 8-16 in saccate, fleshy involucres, on lower surface of 6-8-lobed disciform receptacle with marginal sinuses dorsally, raised on stalk with two rhizoidal furrows; after fertilization and maturation, each involucre generally containing a single sporophyte consisting of foot, seta and capsule; capsule wall unistratose, with annular thickenings, dehiscing irregularly. Spores small, papillose. Elaters slender, tapering, 1-3-spirate. Gemmae absent.

Dumortiera hirsuta (Swartz) Nees in Reinwardt, Blume & Nees ab Esenbeck, Nova Acta Academiae Caesareae Leopoldino-Carolinae Germanicae Naturae Curiosorum XII: 410 (1824); Gottsche et al.: 542 (1846); Spruce: 566 (1885); Stephani: 224 (1899); Macvicar: 41 (1926); Sim: 25 (1926); Müller: 396 (1951); S. Arnell: 52 (1963); Hässel de Menéndez: 182 (1963). Type: Jamaica, leg. Swartz s.n. [S, holo.!; MW, iso. (Hb. Hoffm. No. 8497); UPS, fide Grolle 1976].

Marchantia hirsuta Swartz: 145 (1788).

M. irrigua Wilson ex Hooker in Smith: 106 (1833). Hygropyla irrigua (Wilson) Taylor: 390 (1836); Mackay: 54 (1836); Stephani: 150 (1899). Dumortiera irrigua (Wilson) Nees: 159 (1838). D. hirsuta var. irrigua (Taylor) Spruce: 566 (1885). Type: Ireland, Turk Cascade, near Killarney, Mr Wilson.

- D. hirsuta var. angustior Gottsche, Lindenberg & Nees: 544 (1846).
- D. hirsuta var. intermedia Gottsche, Lindenberg & Nees: 544 (1846).
- D. velutina Schiffner: 256 (1893).

Askepos brevipes Griffith: 340 (1849). Type: India, in sylvis umbrosis Tingrei agri, II 1836.

Thallus hygrophyllous, large and creeping, broadly and rather irregularly strap-shaped, uniformly dark green, translucent, thin and flat, but often marginally undulate, wet (Figure 1A); crisped and shrivelled, dull, unable to survive dry; in crowded, overlying patches, once to several times dichotomously or occasionally laterally furcate or

^{*} National Botanical Institute, Private Bag X101, Pretoria 0001. MS. received: 1992-03-31.



FIGURE 1. — Dumortiera hirsuta. A, dorsal view of thallus; B, ventral view of thallus; C, male plant with disciform receptacles at apex; D, young female receptacle seen from above; E, young female receptacle seen from below; F, transverse section of thallus; G, margin of thallus with hairs; H, transverse section of thallus, much enlarged; I, lower cells of costa and vestigial scales in transverse section; J, filiform scale from top of stalk; K, transverse section of stalk with two rhizoidal furrows; L, capsule wall cells with thickenings; M, transverse section of capsule wall. A, F–H, S.M. Perold 2694; B, S.M. Perold 2634; C, Nicholas II76; D, E, I, H. Anderson CH I3495; J–L, Doidge CH 3581. Scale bars: A–C = 2 mm; D–F, J = 1 mm; G, K = 100 μm; H, I, L, M = 50 μm. Illustrations by G. Condy.

with apical innovations; branches $50-95 \times 8-13(-22)$ mm and $\pm 500 \ \mu$ m thick over median, ventrally bulging midrib (also visible from above), laterally gradually thinning out into wide wings (Figure 1F); apex shortly emarginate; margins undulate, occasionally scalloped, sparsely hirsute, hairs $\pm 250.0 \ \mu$ m long, $\pm 12.5 \ \mu$ m wide at base, tapering slightly to somewhat blunt tip (Figure 1G).

Epidermis, pores and air chambers absent; dorsal cells from above, 4-6-sided, variable in shape and size, 27-57 \times (17-)22-32 μ m, in cross section \pm 15 μ m thick, containing chloroplasts; marginal cells thin-walled, longor short-rectangular, $25-62 \times 12-20 \mu m$; midrib with \pm 18 rows of cells of which upper 3 rows larger, rounded, up to 100 \times 137 μ m, with somewhat wavy walls and scattered chloroplasts, lower group of cells angular, smaller, \pm 30 μ m wide, lacking chloroplasts; wings medianly with 3 rows of large cells, decreasing in number to one row at margin, covered by dorsal and ventral layer of smaller chlorophyllose cells; oil bodies quite rare, for the most part confined to scattered cells in the midrib, yellow-brown, elongate or round, $\pm 40 \ \mu m$ across; rhizoids below midrib numerous, mostly smooth, $15-20 \,\mu m$ wide, occasionally tuberculate, 5 µm wide, also in widely spaced, arching strands below wings, radiating toward margins and appressed to ventral face of wings (Figure 1B) or scattered; ventral scales hyaline, vestigial (Figure 11), without appendages, evanescent, only near apex.

Monoicous or dioicous. Antheridiophore subsessile, disciform, 2.75 mm in diameter, 0.6 mm thick in centre, flattening toward sides, containing antheridia, $470 \times 250 \,\mu\text{m}$, ± ovate, acropetally arranged, but not in radiating rows, sunken into disc, borne singly at apical notch of thallus (Figures IC: 2A) on very short, 1 mm diameter, bifurrowed stalk, with rhizoids lining furrows; hyaline, filiform scales on ventral face of disc, the latter encircled by dense outer fringe of bristle-like hairs, 600-1100 µm long, base 20-25 µm wide, a few scattered hairs dorsally. Archegoniophore disciform, 3.75 mm in diameter; dorsal face (Figure 1D) becoming radially grooved by sinuses; ventral face 6-8-lobed (Figure 1E) in radiating rows and then almost star-shaped, sparingly bristled; archegonia in groups of 8-16, enclosed in green, fleshy, saccate involucres, long archegonial necks protruding through narrow slit-like apical openings; receptacle eventually raised on stalk arising at apical notch of thallus between overlapping sides, \pm 40 mm long and \pm 925 μ m wide, cortical cells \pm 15.0 \times 12.5 μ m, inner cells \pm 50 \times 30 μ m; along its length 2 furrows lined with rhizoids (Figure 1K) and its top (where joined to disc) encircled by numerous filiform, hyaline scales (Figure 1), up to 3750 μ m long and 300 μ m wide at base, apex only \pm 3 cells wide, cells mostly $\pm 125 \times 22 \ \mu m$; at maturity generally only one sporangium borne in each involucre, rarely two and quite often none, elliptical, 2.5×1.5 mm, supported on elongating seta and protruding from membranous calyptra, pseudoperianth lacking; capsule wall unistratose (Figure 1M) with annular or semi-annular thickenings (Figure 1L), dehiscing irregularly via longitudinal suture lines and releasing large numbers of elaters and spores (Figure 2B). Spores golden brown, semi-transparent, distal face rounded, proximal face \pm flat to slightly peaked in the centre, triradiate mark indistinct, 25-30 µm across its longer axis, ornamented with numerous nodules or

tubercles, irregular in size and shape (Figure 3A). *Elaters* yellow-brown or orange-brown, mostly with doubly spiral strands (Figure 3B), ends tapering, $225-470 \times 7.5-10.0 \mu$ m in middle and 3.5 μ m wide at tips. *Gemmae* absent.

Chromosome number: n = 9 (Berrie 1960, Bornefeld 1987); n = 18 (Tatuno 1941); n = 27 (Tatuno 1941, Berrie 1958). According to Tatuno (1938, 1939) the 9, 18 and 27 chromosome sets present different races with different distributions and edaphic ecology, but he considers them as belonging to the same species.

Only a few of the specimens examined were fertile; most of these were dioicous, with male and female receptacles borne on separate plants. Monoicous plants were quite rarely found, but have frequently been reported in the literature and even bisexual receptacles are known (Evans 1919).

All southern African specimens have been identified as belonging to D. hirsuta: the dorsal face of the thallus is always ± smooth and lacks papilliform cells or traces of air chambers. D. nepalensis and D. velutina, which our material resembles, are considered to belong to the polymorphic species D. hirsuta (Grolle & Piippo 1984). Arnell (1963) referred to material from southern Africa (Cape Province, Transvaal, Natal) as D. hirsuta var. nepalensis. Schuster (1992) considers D. nepalensis (Taylor) Nees to be a distinct taxon under the name of D. hirsuta subsp. nepalensis (Taylor) Schuster which is not found in Africa. D. hirsuta is subcosmopolitan and is widespread in tropical and temperate regions, generally growing in sheltered, wooded, shaded and damp areas, i.e. it is hygrophilous. In southern Africa it is known from northern and eastern Transvaal, Swaziland, Natal, Zululand and eastern, southern and southwestern Cape (Figure 4). Further northwards in Africa, D. hirsuta is reported from (or has been collected in) Zimbabwe (Best 1990), Zambia [S.M. Perold 2670 (PRE)], East African Mountains (Arnell 1956), East Africa (Bizot et al. 1985), Tanzania, Malawi, Mozambique (Bizot & Pócs 1979), Rwanda and Burundi (Vána et al. 1979).

On the basis of flavonoid data, Campbell et al. (1979) found chemical affinities between Dumortiera and Wiesnerella, suggesting that they belong in a common family, Wiesnerellaceae (Inoue 1976). This was accepted by Grolle (1983), but Schuster (1984) has retained Dumortiera in the Marchantiaceae, creating a new subfamily, Dumortieroideae Schust., for it and designating Dumortiera as the type genus. Dumortiera has also been classified in the Dumortieroideae by Bischler (1988), who regards Dumortiera and Wiesnerella as morphologically remotely related, although having similar flavonoid patterns. The treatment of Schuster (1984) is followed here, even though the phylogenetic position of Dumortiera remains a matter of some controversy, as conceded by Schuster (1984). Supporting the above treatment are terpene studies of the genera by Asakawa et al. (1979, 1980a, 1980b, 1981), who found that Wiesnerella denudata and Conocephalum conicum shared 17 of these terpene compounds, but only had three in common with Dumortiera hirsuta. Luteolin 5-Oglucuronide, the flavonoid shared by Dumortiera and Wiesnerella which was considered diagnostic for the Wiesnerellaceae by Campbell et al. (1979), has since also been



FIGURE 2. — Dumortiera hirsuta. A, male plant with subsessile antheridiophore from above; B, stalked archegoniophore with dehisced sporangia, front one releasing many spores and elaters. A, B, Koekemoer 989, × 9.

reported for *Conocephalum* by Porter (1981). Whittemore (1991) therefore cautions against drawing taxonomic conclusions from a small number of compounds.

SPECIMENS EXAMINED

TRANSVAAL. –2230 (Messina): Entabene, (-CC), Bottomley (PRE); Entabene, (-CC), Schelpe 6020 (BOL); Soutpansberg, Entabene, (-CC), Thomas 856 (PRE). 2329 (Pietersburg): Pietersburg, (-CD), Van Vuuren 1469 (PRE); Haenertsburg, (-DD), Putterill 3604 (PRE). 2330 (Tzaneen): Woodbush For. Res., Magoebaskloof, near stream, (-CC), H. Anderson CH 13495, CH 13499 (PRE); Bosman 3188, (PRE); De Hoek For. Res., Debengeni Falls, on stream bank, beyond foot bridge, (-CC), S.M. Perold 2634 (PRE); Woodbush, locally abundant in deep shade on streambank, (-CC), Schelpe 6070 (BOL). 2331 (Phalaborwa): Letaba, (-DC), Scheepers 984 (PRE). 2430 (Pilgrim's Rest): Farm Cyprus near Ofcolaco, in gorge named 'Terrible Hollow', (-AB), H. Anderson CH 4527 (PRE); Mariepskop Forestry Water Works, on outside of watertank, also under overhang, dense shade, (-DB), Vorster 87C (PRE); Mariepskop, near dam in Klaserie River, montane forest, on rock against bank next to stream, in shade, (-DB), Vorster 572; Mariepskop For.,

Bedford footpath in forest on soil of streambed, full shade, (-DB), Vorster 1398 (PRE); Mariepskop For., Blyde River footpath, on exposed tree root at streamside in forest shade, (-DB), Vorster 1472 (PRE); Mariepskop, Blyde River footpath, hanging from vertical sandstone rocks in forest shade, (-DB), Vorster 1473 (PRE); Mariepskop, Magalieskop Res. For., on damp earth bank, dense shade, (-DB), Vorster 1815 (PRE); Mariepskop, (-DB), Van der Schijff 4482 (PRE); Mariepskop, Klaserie River by dam, beneath rocks, (-DB), Van der Schijff 6291 (PRE); Mount Sheba Nat. Res., at the 'Grotto', on dripping cliffs, forming extended mats on wet vertical rock face, (-DC), Jacobsen 4421 (PRE); Mount Sheba Nature Reserve at the 'Grotto', on dripping rock cliffs, (-DC), Perold & Koekemoer 2864 (PRE); Pilgrim's Rest, (-DD), Van der Schiff 6367 (PRE). 2527 (Rustenburg): Nature Res. Cederbergkloof, near Utopia, (-CA), Koekemoer 972 (PRE). 2530 (Lydenburg): Farm Klipsteen, between Lydenburg and Dullstroom, at waterfall, on rock, (-AB), H. Anderson CH 13446 p.p. (PRE); Sabie, Lone Creek Falls, on soil near footpath, (-BA), S.M. Perold 2694 (PRE); Sabie Gorge, (-BB), V.A. Wager 21 (PRE); on road to Lydenburg, at turnoff to Witklip, Coromandel Farm, at waterfall, (-AB), Perold & Koekemoer 2839, 2844 (PRE); Nelspruit Dist., Rooiwal, (-BC), Bosman 3180 (PRE); Rosehaugh, (-BD), T.R. Sim CH 1286 (PRE); Kaapsehoop, (-DB), H.A. Wager 47 (PRE); Berlin State Forest, Kaapsehoop hiking trail near Battery Creek, (-DA), Koekemoer 973, 975 (PRE).



FIGURE 3. — Dumortiera hirsuta. A, spore, distal view; B, part of elater. A, B, H. Anderson CH 4527. A, \times 1730; B, \times 480. SEM micrographs by S.M. Perold.

SWAZILAND. - 2531 (Komatipoort): King Forest, Havelock, occasionally on wet shaded earth bank in forest, (-CC), *Schelpe 6195* (BOL).

NATAL. - 2731 (Louwsburg): Ngoma For., bank of river, (-CD), Gerstner 4386 (PRE); Ngoma For., Cetshwayo waterfall walk, vertical slope, seepage, in deep shade, (-CD), Glen 2881 (PRE); Ngoma For., along path in Ntendeka, alongside streams in forest interior, (-CD), Nicholas 1176 (PRE); Ngoma Forest, occasionally on deeply shaded earth banks in forest, (-CD), Schelpe 6237 (BOL); Ngoma For., (-CD), T.R. Sim CH 1302 (PRE); Ngoma For., Ntendeka, in forest, (-CD), A.E. van Wyk 6973 (PRE). 2828 (Bethlehem): Mont-aux-sources, (-DD), Doidge 171 (BOL, PRE). 2831 (Nkandla): Nkandla Forest, common on rocks in and near water in shade, (-CA), Nixon 57 (BOL); Eshowe, Signal Hill, (-CD), Van der Plank CH 1290, CH 1298 (PRE); Ngoya, (-DC), T.R. Sim CH 1289 (PRE). 2929 (Underberg): Mpendle Dist., 6 miles along Everglades/Boston Road River on stream bank, in shade, (-DB), Moll 726 (BOL, PRE); Home Rule, Polela, (-DC), T.R. Sim CH 1305 (PRE); Xumeni For., (-DD), Doidge 3581 (PRE); Donnybrook, (-DD), Scott 3756 (PRE). 2930 (Pietermaritzburg): Karkloof, (-AC), T.R. Sim CH 1291 (PRE); Buccleugh, (-AD), T.R. Sim CH 1301 (PRE); Pietermaritzburg, Town Bush, (-CB), T.R. Sim 7523, 7534; Waterfall, (-CB), T.R. Sim 7548, 7574, 7586 (PRE); Pietermaritzburg, (-CB), T.R. Sim 1296 (PRE); Sweetwater stream, (-CB), T.R. Sim 1300 (PRE); Zwaartkop, (-CB), T.R. Sim CH 1328 (PRE).

CAPE. --3226 (Fort Beaufort): Katberg For., (-BC), Garabedian S.A.M.H. 49743 (BOL; PRE); Hogsback, (-DB), T.R. Sim 1895; Van der Bijl 176; Young 1303 (PRE). 3227 (Stutterheim): Perie Forest, (-CC), T.R. Sim 7534 (BOL). 3318 (Cape Town): Cape Town, (-CD), H.A. Wager 28 (PRE). 3322 (Oudtshoorn): Rust en Vrede, at Oudtshoorn, (-CA), ex Herb. C. vanden Berghen (BOL). 3323 (Willowmore): Gouna Forest, near Lily vlei, (-CC), S. Arnell 1736 (BOL); Bloukrans River Pass, at bridge in forest, on rocks along stream, (-DC), Stirton 9648 (PRE); Bloukrans, along pass in river ravine, on vertical cliff face, (-DC), Zantovska 155 (PRE). 3419 (Caledon): Oudebos, Rivier-sonder-

LUNULARIA Adanson

Lunularia Adanson, Familles des plantes 2: 15 (1763); Micheli: 4 (1729); Nees ab Esenbeck: 29 (1838); Gottsche et al.: 510 (1846); Schiffner: 35 (1893); Stephani: 216 (1899); Howe: 59 (1899); Macvicar: 38 (1926); Sim 123 (1926); Müller: 366 (1951–1958); S. Arnell: 73 (1963); Hässel de Menéndez: 125 (1963). Type species: Lunularia cruciata (L.) Dum. ex Lindb.

Selenia J. Hill: 120 (1773) nom. illeg. Staurophora Willd. 3: 101 (1809). Dichominum Neck.: 345 (1790) (as subgenus). Marsilia O. Kuntze 2: 837 (1891). Sedgwickia S. Bowdich: 35 (1825).

Thallus large, flat, somewhat glossy, green, in overlying patches, sometimes in extensive turfs; on damp soil in old gardens, nurseries and forested areas, may have been introduced into southern Africa. Branches dichotomously furcate toward apex, new growth by apical or lateral innovations; thickened over midrib, gradually thinning toward slightly undulate, hyaline margins; apex emarginate. Dorsal epidermis persistent, hyaline, cell walls sometimes thickened at corners or entirely; air pores simple, elevated and conspicuous, surrounded by several concentric rings of cells, leading below into individual air chambers, these in one layer and floored by chlorophyllose layer of 3-5-celled erect, branched filaments; storage tissue compact, cells colourless, sometimes with pitted walls; scattered cells throughout with single, large, brown oil body; rhizoids numerous, between ventral scales and on midrib, some smooth, others tuberculate; scales ventral, in curved parallel rows on either side of midrib, with round reniform appendages, several cells conor taining oil bodies.

Dioicous. Antheridia sunken into slightly elevated, oval or kidney-shaped, disciform receptacles encircled by raised, membranous sheath, at sides of male thalli (although originally terminal). Archegonia enclosed in terminal receptacles, but by continued growth of thallus



FIGURE 4.—Distribution of *Dumortiera hirsuta*, ●; and *Lunularia cruciata*, □, in southern Africa.



FIGURE 5. — Lunularia cruciata. A, dorsal view of thallus with gemma cups; B, female plant with young archegoniophores; C, ventral view of thallus with new lateral branch; D, transverse section of thallus; E, transverse section of midrib region, much enlarged; F, transverse section of air chamber; G, air pore seen from above; H, margin of thallus with hyaline cells, seen from above; I, older scale; J, young scale; K, longitudinal section through young archegoniophore; L, longitudinal section through gemma. A, D–H, J, L, M, S. M. Perold 2821; B, C, S.M. Perold 1996. Scale bars: A–C = 2 mm; D, E, K, L = 1 mm; I, J = 500 μm; F, M = 100 μm; G, H = 50 μm. Illustrations by A. Pienaar. Figure 1K partly after Benson-Evans & Hughes fig. 4.

Bothalia 23,1 (1993)

laterally situated, conical, white and bud-like when young, sheathed in layers of scales and slightly sunken into rounded depression with distinct rim; after fertilization and further growth, four tubular involucres in the form of a cross are formed, each enclosing one or two sporophytes, composed of foot, seta and capsule, raised on unfurrowed, hairy stalk; capsules eventually exposed by elongation of seta, the wall unistratose, lacking annular thickenings, dehiscing by 4 valves. *Spores* very small, green or brown, smooth. *Elaters* long, tapering and bispiral. *Gemmae* numerous, disc-shaped, inside crescentshaped ridge; wholly diagnostic. *Sporophyte* virtually unknown in southern Africa.

Lunularia cruciata (L.) Dum. ex Lindberg, Notiser Sällskap pro Fauna et Flora Fennica Förhandlingar 9: 298 (1868); Howe: 60 (1899); Macvicar: 40 (1926); Sim: 24 (1926); Müller: 366 (1951–1958); S. Arnell: 73 (1963); Hässel de Menéndez: 126 (1963); E.O. Campbell: 31 (1965). Type: In Europae umbrosis [OXF, syn.; H-SOL, isosyn., fide Grolle (1976)]. For detailed synonymy see K. Müller (1951–1958).

Thallus moderately large and flat, ribbon-like or margins somewhat irregular (Figure 5A), glossy, bright green to yellowish green, with outlines of subdorsal air chambers faintly visible from above, each of the polygonal areas with a central air pore, when wet; reticulum indistinct, leathery, when dry; in crowded overlying patches, dichotomously or irregularly furcate or with apical or lateral innovations from ventral side of thallus (Figure 5C). Branches 40–55 \times 5–8(–10) mm, \pm 650(–1000) μ m thick over median, ventrally bulging midrib, gradually thinning out laterally into wide wings (Figure 5D); apex emarginate or sinusoidal; margins slightly undulate and somewhat scalloped, with outer 4(5) cell rows hyaline.

Dorsal epidermal cells hyaline, in one layer, 5- or 6-sided to irregular in shape, $35-50 \times 20-30 \ \mu m$, thinwalled to somewhat thicker-walled or only thickened at corners, in cross section $20-25 \mu m$ thick; marginal cells with outermost row short- to long-rectangular (Figure 5H), $15-27 \times 10-12 \ \mu m$, cells of inner rows polygonal, 17-27 \times 25 µm; air pores simple, oval, raised, 17–25 \times 12–20 μ m in diameter, bordered by 3–5 rings of curved, smaller, thin-walled cells (Figure 5G), $7-12 \times 17-27 \ \mu m$, outer row of cells somewhat larger, $15-20 \times 20-22 \ \mu m$; air chambers with domed roof (Figure 5F) raised $37-52 \ \mu m$ above filaments, laterally separated by non-chlorophyllose unistratose partitions obscured by filaments, floored by dense chlorophyllose layer, \pm 70 μ m thick, of 3-5-celled erect, branched filaments, filled with chloroplasts, top cell often clavate, $\pm 20 \times 15 \ \mu m$, others $20 \times 12 \ \mu m$; midrib below assimilation cells, with \pm 15(-20) rows of compact colourless storage cells (Figure 5E), $50-75 \times 45-50$ μ m, becoming smaller ventrally, some with pitted walls, layers gradually decreasing in the wings; scattered cells with brown oil bodies, round or oval, $30 \times 30-45 \times 27$ μ m; ventral epidermal cells 30-40 μ m wide, 15-25 μ m thick in cross section; rhizoids on midrib between scales, numerous, smooth, \pm 30 μ m wide, tuberculate, (10-) $17-22 \ \mu m$ wide. Scales hyaline, some basal cells purple, on either side of midrib, stretched across ventral face of wings, near apex of thallus, base $\pm 1375 \times 550 \,\mu\text{m}$, with rounded, constricted appendage (Figure 5J), $\pm 400 \times 250$ μ m; cells mostly polygonal, $\pm 65 \times 25 \mu$ m, some smaller, with oil body 22 $\times 25 \mu$ m, almost entirely filling cell; older scales larger (Figure 5I), base up to 1000 \times 4250 μ m, appendage $\pm 600 \mu$ m wide.

Dioicous. Male plants quite rare in southern Africa. Antheridiophore on alternate sides, having originally developed in terminal sinus near apex (Figure 6A), slightly raised, ovate, flattish discs, 2-3 mm in diameter, encircled by membranous sheath with crenate edges, containing numerous antheridia, individually sunken in flask-shaped cavities (Figure 6B), opening above by pores. Archegoniophore originally also developing in terminal sinus near apex, with further growth of thallus leaving it behind, so that it appears lateral in position on alternate sides of thallus (Figure 5B); commences as domed disc \pm 200 \times 375 µm, bearing several archegonia in radiating rows and attached by very short stalk to floor of shallow, round hollow, 1100 μ m wide \times 300 μ m deep, with crenate rim; sheathed in \pm 3 layers of scales (Figure 5K), outer layer formed by fused, shaped scales, bulging in centre and constricted below and above, upper edge irregularly fringed with filiform cellular appendages of up to 7 rectangular cells, \pm 35 × 15 μ m, cells in body of scale thick-walled, 5- or 6-sided, up to 57 \times 25 μ m, in between numerous smaller cells, 4- or 5-sided, $25 \times 15 \mu m$, almost entirely filled with single oil body; between scales and especially from base of archegoniophore and arching over it, numerous uniseriate, long hairs of 16-20 cells each. Further development not recorded as fertilization did not take place, nor are sporophytes available for study. Gemmae numerous, disc-shaped (Figure 5M), notched



FIGURE 6. — Lunularia cruciata. A, antheridiophore seen from above; B, longitudinal section through antheridiophore, with one flaskshaped antheridium in place. A, Wilman BOL. No. 24870; B, Koekemoer 1004. A, × 22; B, × 38. A, SEM micrograph; B, LM photograph.

by 2 opposite, lateral growing points, $\pm 430 \ \mu m$ in diameter when mature, developing upright on short stalk, inside crescent-shaped cupule (Figure 5L), $\pm 3 \ mm$ wide with crenate to entire ridge on proximal side. *Chromosome number* n = 8 (Heitz 1927); n = 9 = 8 + x/y² (Lorbeer 1934); n = 9 (Bornefeld 1987).

DISCUSSION

As mentioned in the description, male plants with antheridial receptacles are exceedingly rare in southern Africa and the only two, Wilman BOL No. 24870, on loan from BOL and Koekemoer 1004 (PRE), were received after completion of Figure 5 and therefore not illustrated there (see Figure 6). Male plants are so rarely seen, that Sim (1926) had categorically stated that they are not present in South Africa, but then added 'so far as is known'. He must have been unaware of Saxton's find. No female plants with fertilized archegonia and mature sporophytes were available for study; even plants with young archegoniophores are quite rare, judging both from personal experience and from the literature (Saxton 1931; Goodman 1956; E.O. Campbell 1965). Saxton (1931) found both male and female plants in Cape Town in 1908, but had to wait for almost 20 years for plants with mature archegoniophores (forwarded from Dartmouth, England) to complete his studies on the life history of L. cruciata. Giffen sent plants with young archegoniophores from Oranjezicht, Cape Town to Sim (Sim 1926) and Auret (as mentioned in Benson-Evans & Hughes 1954) reported the regular production of female branches in the vicinity of Johannesburg. Of my own collections, only S.M. Perold 1996 from Devon Valley Hotel, Stellenbosch (November 1987), had young archegoniophores.

Since so many collections are from nurseries or city gardens, it would appear that L. cruciata may have been introduced into southern Africa. It is not frequently found here, most collections being from the southwestern Cape, a few from southern Transvaal and some from Natal, and then quite frequently from nurseries (Figure 4). Further north in Africa, L. cruciata is known from Zimbabwe (Best 1990), Malawi (Nyika Plateau, S.M. Perold 2667, 2676 (PRE)); East African Mountains (Arnell 1956); Tanzania (Serengeti) (Vanden Berghen 1965); Uluguru Mountains, Rungwe Mountains (Bizot & Pócs 1979); Congo Rep. (Katanga) (Vanden Berghen 1965). Pócs (pers. comm.) states that in East Africa, L. cruciata is found in many natural habitats in the montane forest belt, mostly on young volcanoes such as Mt Elgon and Mt Meru and even on the Comoro Islands.

Lunularia cruciata is quite widespread in the southern hemisphere but regarded by Engel & Schuster (1982) as probably Laurasian in origin. Since sexual reproduction and subsequent spore production are so rare, its gemmae obviously present a highly effective means of ensuring its dispersal, which is most likely aided by human activities.

On the basis of its flavonoid chemistry, Campbell *et al.* (1979), include it in the Marchantiaceae, but in the present treatment Grolle (1983) is followed and *L. cruciata* is classified in the monotypic family, Lunulariaceae Klinggr. (1858).

Its phylogenetic position seems rather unclear, Schuster (1984b) arguing that he would place it 'low' in the Marchantiales on account of its high level of seta retention, 2-3 sporophytes per gynoecium and a capsule with 4 well-defined valves. On the other hand, he expresses the viewpoint (Schuster 1984b) that its archegoniophore, clearly formed from two dichotomies, each producing archegonia, is complex and therefore an advanced feature.

SPECIMENS EXAMINED

TRANSVAAL. –2528 (Pretoria): Pretoria, Union Buildings Nurseries, (-CA), Bottomley CH 135 (PRE); Pretoria, National Botanical Institute Nurseries, (-CA), S.M. Perold 2821 (PRE). 2627 (Potchefstroom), Vereeniging, (-DB), T.R. Sim CH 1283 (PRE); Roodepoort, Sterlig Nursery, on gravel and brick walls of flowerbeds, (-DD), Koekemoer 1004 (PRE).

NATAL. –2930 (Pietermaritzburg): Hilton Road, (-CB), T.R. Sim CH 1279, CH 1280 (PRE). 2931 (Stanger): Durban, Silverglen Nurseries, (- CC), S.M. Perold 2805 (PRE).

CAPE. - 3318 (Cape Town): Kirstenbosch, (-CD), S. Arnell 412 (BOL); Window Gorge, Table Mountain, (-CD), S. Arnell 405 (BOL); Claremont Park, (-CD), Garside 6653 (BOL); near Round House, (-CD), Garside 6132 (BOL); Oranjezicht, (-CD), Giffen CH 1281 (PRE); Glen Picnic Resort, just below Round House, Lion's Head, (-CD), S.M. Perold 645 (PRE); Round House, Lion's Head, on soil beneath trees, (-CD), S.M. Perold 650 (PRE); Newlands Forest, on soil under trees, (-CD), S.M. Perold 662 (PRE); Cape Town, (-CD), T.R. Sim CH 1278, CH 1282 (PRE); Skeleton Gorge, rock face, very wet forest, (-CD), Stirton 9415 (PRE); mountain slopes above Kirstenbosch, (-CD), Stokoe s.n. (BOL); Kirstenbosch, (-CD), Wilman 24870 (BOL); Stellenbosch, (-DD), Burtt Davy CH 1277 (PRE); Pniel, on tree trunk mixed with moss, (-DD), Morley 314 (PRE); Stellenbosch, Devon Valley Hotel at garden fence, on soil, (-DD), S.M. Perold 1990 (PRE); Stellenbosch, Devon Valley Hotel, on soil bank behind hotel, (-DD), S.M. Perold 1996 (PRE); Stellenbosch, (-DD), T.R. Sim CH 1284 (PRE); S Paarl, Landskroon, in kloof, (-DD), Volk 81/071 (BOL, PRE). 3319 (Worcester): W of Franschhoek, Waterval Farm, on earth bank, ditch next to dirt road, (-CC), S.M. Perold 633 (PRE); 4 km N of Villiersdorp, Elandsriver Road, Du Toitsberge, near waterfall on Sneeukop, (-CD), S.M. Perold 623 (PRE). 3322 (Oudtshoorn): George, NE of Hawthorndene Hotel, at roadside on side of earth water furrow, (-CD), S.M. Perold 920 (PRE). 3418 (Simonstown): Constantia, on soil on damp rock face, (-AB), S.M. Perold 656 (PRE).

ACKNOWLEDGEMENTS

I wish to express my gratitude to Dr R. Grolle for his information regarding the publication date of 'Hepaticae Javanicae' by Reinwardt *et al.*, as well as for drawing my attention to Nelson & Parnell's publication on the synonymy of the various forms of *Hygropyla* etc. under *Dumortiera*. I also thank Dr Grolle, Prof. T. Pócs and Prof. O.H. Volk for critically reading the manuscript and for their valuable suggestions. Further thanks are due to the curator of BOL for the loan of specimens; to the artists, Ms G. Condy and Ms A. Pienaar; to the typist, Mrs J. Mulvenna and to the photographer, Mrs A. Romanowski. Also to my colleagues at NBI, particularly Drs H. Anderson and H.F. Glen, as well as Miss M. Koekemoer for collecting specimens.

REFERENCES

- ADANSON, M. 1763. Familles des plantes 2. Paris.
- ARNELL, S. 1956. Hepaticae collected by O. Hedberg et al. on the East African mountains. Arkiv für Botanik 3: 517–562.
- ARNELL, S. 1963. Hepaticae of South Africa. Swedish Natural Science Council, Stockholm.

Bothalia 23,1 (1993)

- ASAKAWA, Y., TOKUNAGA, N., TOYOTA, M., TAKEMOTO, T. & SUIRE, C. 1979. Chemosystematics of bryophytes I. The distribution of terpenoids in bryophytes. *Journal of the Hattori Botanical Laboratory* 45: 395-407.
- ASAKAWA, Y., MATSUDA, R. & TAKEMOTO, T. 1980a. Mono- and sesquiterpenoids from Wiesnerella denudata. Phytochemistry 19: 567-569.
- ASAKAWA, Y., TOKUNAGA, N., TAKEMOTO, F., HATTORI, S., MIZUTANI, M. & SUIRE, C. 1980b. Chemosystematics of bryophytes IV. The distribution of terpenoids and aromatic compounds in Hepaticae and Anthocerotae. Journal of the Hattori Botanical Laboratory 47: 153-164.
- ASAKAWA, Y., MATSUDA, R. & TAKEDA, R. 1981. Mono- and sesquiterpenoids of *Conocephalum supradecompositum*. *Phytochemistry* 20: 1423, 1424.
- BENSON-EVANS, K. & HUGHES, J.G. 1954. The physiology of sexual reproduction in *Lunularia cruciata* (L.) Dum. *Transactions of* the British Bryological Society 2: 513-521.
- BERRIE, G.K. 1960. The chromosome numbers of liverworts (Hepaticae and Anthocerotae). *Transactions of the British Bryological Society* 3: 688-705.
- BEST, E.B. 1990. The Bryophyta of Zimbabwe—an annotated checklist. Kirkia 13: 293–318.
- BISCHLER, H. 1988. Relationships in the order Marchantiales (Hepaticae). Journal of the Hattori Botanical Laboratory 64: 47-57.
- BIZOT, M. & PÓCS, T. 1979. East African bryophytes, III. Acta Botanica Academiae Scientiarum Hungaricae 25: 223–261.
- BIZOT, M., PÓCS, T. & SHARP, A.J. 1985. Results of a bryogeographical expedition to East Africa in 1968, III. *The Bryologist* 88: 135-142.
- BORNEFELD, T. 1987. The natural system of the Marchantiales based upon cytogenetical and morphological evidence. *Nova Hedwigia* 45: 41–52.
- BOWDICH, S. 1825. Excursions in Madeira and Porto Santo: 35.
- CAMPBELL, E.O. 1965. Lunularia in New Zealand. Tuatara 13: 31-42.
- CAMPBELL, E.O., MARKHAM, K.R., MOORE, N.A., PORTER, L.J. & WALLACE, J.W. 1979. Taxonomic and phylogenetic implications of comparative flavonoid chemistry of species in the family Marchantiaceae. Journal of the Hattori Botanical Laboratory 45: 185-199.
- ENGEL, J.J. & SCHUSTER, R.M. 1982. Austral Hepaticae XV. Brevianthaceae: a monotypic family endemic to Tasmania. The Bryologist 85: 375-388.
- EVANS, A.W. 1919. A taxonomic study of Dumortiera. Bulletin of the Torrey Botanical Club 46: 167–182.
- GOODMAN, G.T. 1956. Sexual Lunularia cruciata (L.) Dum. in South Wales. Transactions of the British Bryological Society 3: 98-102.
- GOTTSCHE, C.M., LINDENBERG, J.B.G. & NEES AB ESENBECK, C.G. 1844–1847. Synopsis Hepaticarum. Hamburg.
- GRIFFITH, W. 1849. On the higher cryptogamous plants. Notulae ad plantas asiaticas, part 2: 285-352. Calcutta.
- GROLLE, R. 1976. Verzeichnis der Lebermoose Europas und benachbarter Gebiete. Feddes Repertorium 87: 171-279.
- GROLLE, R. 1983. Nomina generica Hepaticarum, references, types and synonymies. Acta Botanica Fennica 121: 1-62.
- GROLLE, R. & PIIPPO, S. 1984. Annotated catalogue of western Melanesian bryophytes 1. Acta Botanica Fennica 125: 1–86.
- HÄSSEL DE MENÉNDEZ, G.G. 1963. Estudio de las Anthocerotales y Marchantiales de la Argentina. Opera Lilloana 7: 1–297.
- HEITZ, E. 1927. Über multiple und aberrante Chromosomenzahlen. III. Das Genom der Lebermoose. Abhandlungen der naturwissenschaftlichen Verein zu Hamburg 21: 48-58.
- HILL, J. 1773. A general natural history 2, edn 2: 120.
- HOOKER, J.D. & TAYLOR, T. 1844. Flora antarctica. London Journal of Botany 3: 576.
- HOWE, M.A. 1899. The Hepaticae and Anthocerotes of California. Memoirs of the Torrey Botanical Club 7: 13-33.

- INOUE, H. 1976. Illustrations of Japanese Hepaticae 2. Tokyo.
- KLINGGRAEFF, H.E.M. 1858. Die höheren Cryptogamen Preussens. Königsberg.
- KUNTZE, O. 1891. Revisio generum plantarum 2: 837.
- LINDBERG, S.O. 1868. Musci novi scandinavici. Notiser Sällskap pro Fauna et Flora Fennica Förhandlingar 9: 298.
- LORBEER, G. 1934. Die Zytologie der Lebermoose mit besonderer Berücksichtigung allgemeiner Chromosomenfragen. Jahrbuch für wissenschaftliche Botanik 80: 567-818.
- MACKAY, J.T. 1836. Flora hibernica. Dublin.
- MACVICAR, S.M. 1926. The student's handbook of British hepatics: 464. Eastbourne.
- MICHELI, P.A. 1729. Nova plantarum genera. Florintiae.
- MÜLLER, K. 1951–1958. Die Lebermoose Europas, in Rabenhorst's Kryptogamenflora 6: 416–471.
- NECKER, N.J. DE 1790. Elementa: 345.
- NEES AB ESENBECK, C.G.D. 1824. Dumortiera. In Reinwardt, Blume & Nees ab Esenbeck, Hepaticae javanicae. Nova Acta Academiae Caesareae Leopoldina-Carolinae Germanicae Naturae Curiosorum XII: 410.
- NEES AB ESENBECK, C.G.D. 1838. Naturgeschichte der europäischen Lebermoose 4. Berlin & Breslau.
- NELSON, E.C. & PARNELL, J. 1992. Flora hibernica (1836): its publication and aftermath as viewed by Dr Thomas Taylor. Taxon 41: 35-42.
- PORTER, L.J. 1981. Geographic races of *Conocephalum* (Marchantiales) as defined by flavonoid chemistry. *Taxon* 30: 739-748.
- RADDI, G. 1818. Novarum vel rariorum ex Cryptogamia stirpium in agro Florentino collectarum Decades duae. *Opuscoli scientifici* di Bologna 2: 349-361.
- SAXTON, W.T. 1931. The life history of *Lunularia cruciata* (L.) Dum., with special reference to the archegoniophore and sporophyte. *Transactions of the Royal Society of South Africa* 19: 259-268.
- SCHIFFNER, V.F. 1893. Hepaticae bearbeitet von Dr Victor Schiffner, aus Engler-Prantl. Die natürlichen Pflanzenfamilien 1: 3-141.
- SCHUSTER, R.M. 1984a. Diagnoses of some new taxa of Hepaticae. *Phytologia* 56: 65-74.
- SCHUSTER, R.M. 1984b. New manual of bryology 2: 892-1070. The Hattori Botanical Laboratory, Nichinan, Miyazaki, Japan.
- SCHUSTER, R.M. 1992. The Hepaticae and Anthocerotae of North America. Vol. 6.
- SIM, T.R. 1926. The Bryophyta of South Africa. Transactions of the Royal Society of South Africa 15: 1-475.
- SMITH, J.E. 1833. Cryptogamia. The English Flora 5: 106.
- SPRUCE, R. 1855. Hepaticae amazonicae et andinae. Transactions of the Botanical Society, Edinburgh 15: 1-590.
- STEPHANI, F. 1899. Species hepaticarum. Bulletin de l'Herbier Boissier 7: 198–225.
- SWARTZ, O. 1788. Nova genera et species plantarum: 145.
- TATUNO, S. 1938. Über Polyploidie und geographische Verbreitung bei Dumortiera hirsuta. Botanical Magazine Tokyo 52: 434-441.
- TATUNO, S. 1939. Weitere Untersuchungen über die Polyploidie und die geographische Verbreitung bei Dumortiera hirsuta. 1–11. Botanical Magazine Tokyo 53: 345–350.
- TATUNO, S. 1941. Zytologische Untersuchungen über die Lebermoose von Japan. Journal Scientifica Hiroshima University 4: 73-187.
- TAYLOR, T. 1836. De Marchantieis. *Transactions of the Linnean Society* 17: 375–395.
- VÁŇA, J., PÓCS, T. & DE SLOOVER, J.L. 1979. Hepatiques d'Afrique tropicale. Lejeunia 98: 1-23.
- VANDEN BERGHEN, C. 1965. Hépatiques récoltées par le Dr J.-J. Symoens dans la région péri-Tanganyikaise. Bulletin de la Société Royale de botanique de Belgique 98: 129–174.
- WHITTEMORE, A.T. 1991. The secondary chemistry of the Marchantiales. Advances in Bryology 4.
- WILLDENOW, C.L. 1809. Gesellschaft naturforschender Freunde, Berlin. Magazin für die neuesten Entdeckungen 3: 101.